

## Kinetic spectrophotometric analysis of naftidrofuryl oxalate and vincamine in pharmaceutical preparations using alkaline potassium permanganate

Follow this and additional works at: <https://www.jfda-online.com/journal>

### Recommended Citation

Belal, T.S.; Barary, M.H.; Sabry, S.M.; and Ibrahim, M.E.A.-L. (2009) "Kinetic spectrophotometric analysis of naftidrofuryl oxalate and vincamine in pharmaceutical preparations using alkaline potassium permanganate," *Journal of Food and Drug Analysis*: Vol. 17 : Iss. 6 , Article 1.  
Available at: <https://doi.org/10.38212/2224-6614.2575>

This Original Article is brought to you for free and open access by Journal of Food and Drug Analysis. It has been accepted for inclusion in Journal of Food and Drug Analysis by an authorized editor of Journal of Food and Drug Analysis.

# Kinetic Spectrophotometric Analysis of Naftidrofuryl Oxalate and Vincamine in Pharmaceutical Preparations Using Alkaline Potassium Permanganate

TAREK SAIED BELAL<sup>1\*</sup>, MAGDA HAMDY BARARY, SUZY MOHAMMED SABRY  
AND MOHAMMED ELSAYED ABDEL-LATIF IBRAHIM

Pharmaceutical Analytical Chemistry Department, Faculty of Pharmacy, University of Alexandria, Elmessalah 21521, Alexandria, Egypt

(Received: January 14, 2009; Accepted: October 20, 2009)

## ABSTRACT

A simple, rapid and sensitive kinetic spectrophotometric method is described for the analysis of naftidrofuryl oxalate (NF) and vincamine (VN) in pure form and in their pharmaceutical preparations. The procedure was based on the kinetic investigation of the oxidation of the studied drugs with alkaline potassium permanganate and the absorbance of the produced manganate species was measured at 610 nm. Variables affecting the color development were investigated and the conditions were optimized. The concentration of the studied drugs was calculated using the regression equations for the fixed-time method. The determination of the two drugs by fixed absorbance and rate constant methods was possible but the fixed time method was more applicable. The reliability and analytical performance of the proposed method including linearity, ranges, precision, accuracy, detection and quantification limits were statistically validated. Calibration graphs are linear over the concentration ranges of 3-15 µg/mL and 4-14 µg/mL for NF and VN, respectively. The proposed method was satisfactorily applied for analysis of pharmaceutical preparations containing the studied drugs. A proposal of the reaction pathway was postulated.

Key words: naftidrofuryl oxalate, vincamine, potassium permanganate, kinetic spectrophotometry, pharmaceutical preparations

## INTRODUCTION

Naftidrofuryl oxalate (NF), 2-(diethylamino)ethyl 2-[(naphthalen-1-yl)methyl]-3-(tetrahydrofuran-2-yl) propanoate hydrogen oxalate, is a vasodilator drug used in the treatment of peripheral and cerebral vascular disorders. It is claimed to enhance the cellular oxidative capacity thereby protecting cells against ischemia<sup>(1)</sup>. NF is an official drug in the British Pharmacopoeia<sup>(2)</sup> where a nonaqueous potentiometric titration and HPLC procedures were described for the assay of NF bulk powder and capsules, respectively. Few analytical methods have been reported for the determination of NF in biological fluids and/or pharmaceutical preparations. Most of these studies focused on HPLC-UV detection<sup>(3,4)</sup>, HPLC-fluorescence detection<sup>(5-8)</sup> and phosphorimetric analysis<sup>(9-12)</sup>. Others included potentiometric method using NF ion-selective electrodes<sup>(13)</sup>, flow injection analysis with

fluorescence optosensor<sup>(14)</sup> and spectrophotometry<sup>(15)</sup>.

Vincamine (VN), methyl (3 $\alpha$ , 16 $\alpha$ )-14, 15-dihydro-14 $\beta$ -hydroxy eburnamenine-14-carboxylate, is an alkaloid obtained from *Vinca minor* (Apocyanaceae). It is claimed to increase cerebral circulation and utilization of oxygen and has been used in a variety of cerebral disorders<sup>(1)</sup>. Several methods have been reported for the determination of VN in different matrices. VN has been determined in biological fluids by HPLC<sup>(16-19)</sup> and GC<sup>(20-23)</sup> methods. A fluorimetric method was described for the estimation of VN in biological fluids obtained from rats<sup>(24)</sup>. In pharmaceutical preparations, VN has been determined by derivative and chemometric spectrophotometry<sup>(25-28)</sup>, HPLC<sup>(19,25,27-29)</sup>, TLC<sup>(28)</sup> and PMR<sup>(30)</sup> procedures. Gravimetric, titrimetric and spectrometric analyses of VN were carried out based on its complexation with thiocyanate ion<sup>(31,32)</sup>. VN has been also determined in plant tissue cultures of *Vinca minor* by TLC<sup>(33)</sup> and HPLC<sup>(34)</sup> methods.

Analytical procedures based on kinetic

\* Author for correspondence. Tel: +20-3-4871317;  
Fax: +20-3-4871351; E-mail: tbelaleg@yahoo.com

spectrophotometry are still lacking in the literature for the determination of NF or VN in pharmaceutical formulations. Kinetic methods are considered of great interest in chemical and pharmaceutical analysis<sup>(35)</sup>. Potassium permanganate in alkaline medium easily reacts with compounds susceptible to oxidation with the formation of manganate species which can be followed up spectrophotometrically. Several pharmaceutical compounds were determined through this approach<sup>(36-40)</sup>. In this work, a kinetics-based spectrophotometric method was developed for the assay of NF and VN. The method is based on oxidizing the two drugs with alkaline  $\text{KMnO}_4$  at room temperature, and subsequently the rate of appearance of the green colored product was monitored at 610 nm. The proposed method is simple, rapid, cost-effective and readily adaptable to both bulk powders and dosage forms, hence more suitable for the application in quality control laboratories in developing countries.

## MATERIALS AND METHODS

### I. Apparatus

Spectrophotometric measurements were performed on a Perkin-Elmer Lambda EZ201 UV-visible spectrophotometer (PerkinElmer, Waltham, Massachusetts,

USA) with matched 1-cm quartz cells.

### II. Drugs and Chemicals

All chemicals and solvents used were of analytical reagent grade. Naftidrofuryl oxalate was kindly donated by Minapharm Pharmaceuticals and Chemical Industries, Cairo, Egypt. Vincamine was kindly provided by GlaxoSmithKline S.A.E., El-Salam city, Cairo, Egypt. Pharmaceutical preparations examined in this study were purchased from the local market and they included Praxilene<sup>®</sup> tablets (Minapharm Pharmaceuticals and Chemical Industries, Cairo, Egypt, under license of Merck Serono, BN. 5AE0028) labeled to contain 200 mg NF per tablet, Oxybral<sup>®</sup> capsules (GlaxoSmithKline S.A.E., El-Salam city, Cairo, Egypt, BN. 071058A) labeled to contain 30 mg VN per capsule and Oxybral<sup>®</sup> ampoules (GlaxoSmithKline S.A.E., El-Salam city, Cairo, Egypt, BN. 074983A) labeled to contain 15 mg VN per 2 mL.

### III. General Procedure

Potassium permanganate solution, 12 mg/mL (0.076 M), and sodium hydroxide solution, 0.5 M, were prepared in distilled water. NF standard solution, 3 mg/mL ( $6.33 \times 10^{-3}$  M), and VN standard solution, 0.5 mg/mL ( $1.41 \times 10^{-3}$  M), were prepared in acetonitrile. Working

**Table 1.** Experimental and analytical parameters for the kinetic determination of NF and VN

Parameter	NF	VN
Temperature (°C)	25 ± 2	25 ± 2
Time (min)	35	20
Concentration range [M]	$6.33 \times 10^{-6}$ - $3.17 \times 10^{-5}$	$1.13 \times 10^{-5}$ - $3.95 \times 10^{-5}$
Molar absorptivity ( $\epsilon$ ) (L mole <sup>-1</sup> cm <sup>-1</sup> )	25795	20593
Regression equation $A = a + b \times C$	$A = 0.0395 + 25794.84 C$	$A = -0.0099 + 20592.51 C$
Correlation coefficient (r)	0.9974	0.9986
$S_a^a$	0.0067	0.0054
$S_b^b$	388.65	200.24
$S_b\%^c$	1.51	0.97
$S_{y/x}^d$	0.0055	0.0047
LOD <sup>e</sup> [M]	$8.01 \times 10^{-7}$	$1.19 \times 10^{-6}$
LOQ <sup>f</sup> [M]	$2.67 \times 10^{-6}$	$3.97 \times 10^{-6}$

<sup>a</sup>: Standard deviation of the intercept.

<sup>b</sup>: Standard deviation of the slope.

<sup>c</sup>: Percentage relative standard deviation of the slope

<sup>d</sup>: Standard deviation of residuals.

<sup>e</sup>: Limit of detection.

<sup>f</sup>: Limit of quantification.

solutions, 300  $\mu\text{g/mL}$  NF ( $6.33 \times 10^{-4}$  M) and 100  $\mu\text{g/mL}$  VN ( $2.82 \times 10^{-4}$  M), were prepared by dilution of standard solutions with acetonitrile. These solutions were found stable for at least 4 days and were stored refrigerated at 4°C during this study.

Accurate volumes of NF or VN working solutions with the concentration ranges shown in Table 1 were transferred into 10-mL volumetric flasks containing the appropriate volumes of potassium permanganate solution (1 mL) and sodium hydroxide solution (3 mL). The solutions were diluted to the volume with distilled water, mixed well and left at room temperature ( $25 \pm 2^\circ\text{C}$ ) for a fixed time of 35 min for NF and 20 min for VN. The absorbance of solutions was measured at 610 nm against similarly treated blanks. The drug concentrations were then computed from the corresponding regression equations of the calibration graphs for the fixed time method.

#### IV. Assay of Pharmaceutical Preparations

##### (I) For NF Tablets

A total of 20 tablets ("Praxilene<sup>®</sup>" tablets) were weighed and finely powdered. To an accurately weighed quantity of the powder containing the equivalent of 30 mg of NF, 60 mL of acetonitrile were added, stirred for 10 min and then filtered into a 100 mL volumetric flask. The residue was washed with two 10 mL portions of acetonitrile, washings were added to the filtrate and diluted to the volume with acetonitrile and aliquots were treated as described in General Procedure.

##### (II) For VN Capsules

To an accurately weighed quantity of the powdered content of Oxybral<sup>®</sup> capsules equivalent to 10 mg of VN, 60 mL of acetonitrile were added and stirred for 10 min and then filtered into a 100 mL volumetric flask. The residue was washed with two 10 mL portions of acetonitrile, washings were added to the filtrate and diluted to volume with acetonitrile and aliquots were treated as described in General Procedure.

##### (III) For VN Ampoules

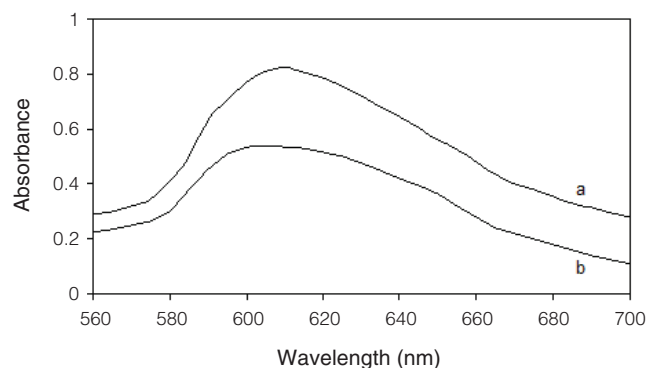
An accurate volume of the mixed contents of five Oxybral<sup>®</sup> ampoules equivalent to 10 mg of VN was transferred into a separating funnel and alkalized with 10 mL of 0.5 M NaOH. VN was extracted by shaking the aqueous layer with 3 portions of chloroform (10 mL each). The organic layer was then collected and evaporated to dryness under vacuum. The residue was dissolved in acetonitrile by sonication for 10 min. It was then transferred quantitatively into a 100 mL volumetric flask, diluted to the volume with acetonitrile and aliquots were treated as described in General Procedure.

## RESULTS AND DISCUSSION

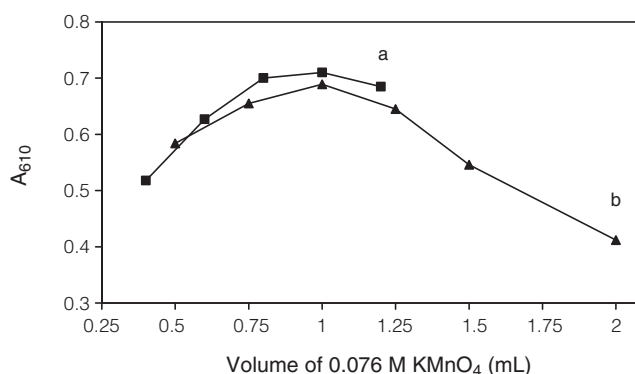
NF and VN react with alkaline potassium permanganate to give the green colored manganate species that absorbs maximally at 610 nm (Figure 1). The extent of formation of this species depends on the concentration of reactants, alkalinity of solution and temperature. Therefore, various experimental parameters affecting the development and the stability of the reaction product were optimized by changing each variable in turn while keeping all others constant.

#### I. Kinetics and Optimization of the Reaction Conditions

The effect of potassium permanganate concentration on the reaction was studied over the range of  $3.04 \times 10^{-3}$  to  $9.12 \times 10^{-3}$  M for NF and  $3.8 \times 10^{-3}$  to  $1.52 \times 10^{-2}$  M for VN. The maximum absorbance was obtained at the concentration  $7.6 \times 10^{-3}$  M (1 mL of 0.076 M  $\text{KMnO}_4$  diluted to final reaction volume 10 mL) for both NF and VN. Higher concentrations of potassium permanganate yielded lower absorbance values, probably due to decomposition of the product (Figure 2). Complete reaction between the investigated drugs and potassium

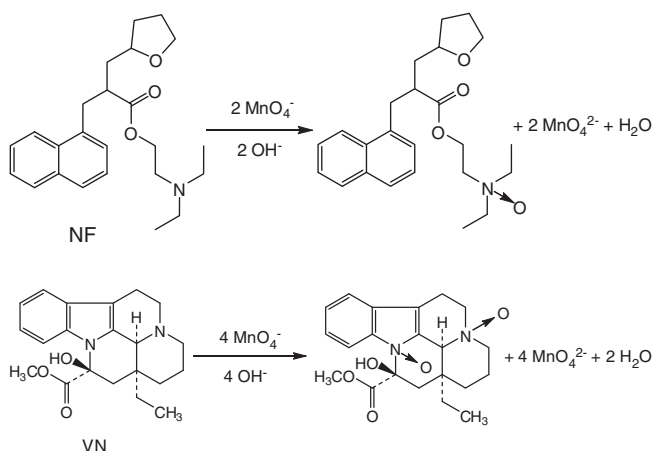


**Figure 1.** Absorption spectra of the reaction product of (a)  $2.53 \times 10^{-5}$  M NF and (b)  $2.82 \times 10^{-5}$  M VN.



**Figure 2.** Effect of  $\text{KMnO}_4$  concentration on the reaction product of (a)  $2.11 \times 10^{-5}$  M NF and (b)  $3.39 \times 10^{-5}$  M VN using optimum conditions for each compound.

permanganate took place only in alkaline medium. The influence of alkalinity was investigated in the range of 0.025 to 0.175 M NaOH for NF and 0.025 to 0.2 M NaOH for VN. The maximum absorbance values were obtained at concentration 0.15 M (3 mL 0.5 M NaOH) for both drugs (Figure 3). The effect of temperature was studied in the range of 25-60°C. The rate of reaction increased with increasing temperature; however,  $25 \pm 2^\circ\text{C}$  was selected as the optimum temperature due to low reproducibility of absorbance values at higher temperature. In order to ascertain the stoichiometry of the studied reactions, Job's method of continuous variation<sup>(41)</sup> was applied. For both drugs, Job's method plot (Figure 4) reached a maximum value at a mole fraction of 0.3 for NF and 0.2 for VN which indicated a reaction ratio of 1: 2 and 1: 4 (drug :  $\text{KMnO}_4$ ) for NF and VN, respectively. The reaction pathway can be explained by the oxidation of the tertiary amine groups into *N*-oxide. It has been reported that compounds possessing tertiary amine groups are liable to oxidation by alkaline potassium permanganate<sup>(39,40)</sup>. The following schemes represent the proposed pathways for the reaction between the investigated drugs and  $\text{KMnO}_4$  in alkaline medium.



The rate of reaction was found to be drug dependant. The rates were followed at room temperature with various concentrations of the investigated drugs in the range of  $6.33 \times 10^{-6}$  to  $3.17 \times 10^{-5}$  M (3-15  $\mu\text{g/mL}$ ) for NF and  $1.13 \times 10^{-5}$  to  $3.95 \times 10^{-5}$  M (4-14  $\mu\text{g/mL}$ ) for VN, keeping  $\text{KMnO}_4$  and NaOH constant at high concentrations as above. The graphs shown in Figures 5 and 6, clearly demonstrated that the reaction rates obey the following equation:

$$\text{Rate} = K' \times [\text{C}]^n \dots\dots\dots (1)$$

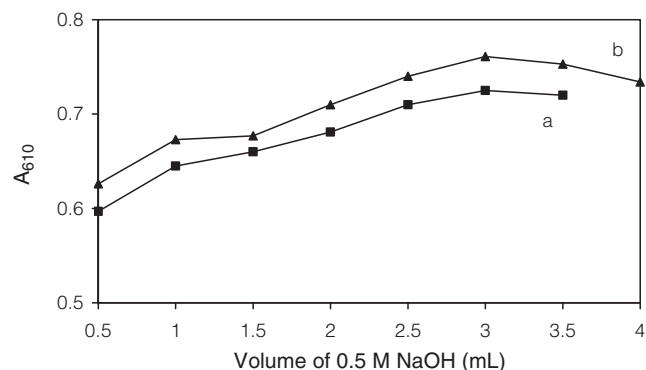
Where  $K'$  is the pseudo-order rate constant and  $n$  is the order of reaction. The rate of reactions could be estimated as  $\Delta A/\Delta t$ , where  $A$  is the absorbance and  $t$  is the time in seconds. Taking logarithms of rates and concentrations (Table 2), equation (1) is transformed into:

$$\log (\text{rate}) = \log (\Delta A/\Delta t) = \log K' + n \log [\text{C}] \dots\dots\dots (2)$$

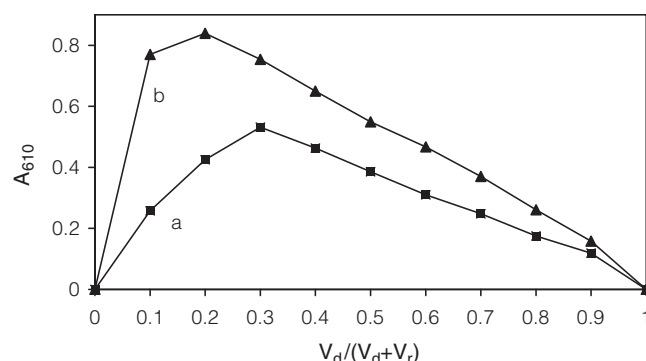
Regression of  $\log (\text{rate})$  versus  $\log [\text{C}]$  gave the following regression equations:

$$\log (\text{rate}) = 0.872603 + 0.89630 \log [\text{NF}], r = 0.998338$$

$$\log (\text{rate}) = 1.382608 + 1.03817 \log [\text{VN}], r = 0.998847$$



**Figure 3.** Effect of NaOH concentration on the reaction product of (a)  $2.11 \times 10^{-5}$  M NF and (b)  $3.39 \times 10^{-5}$  M VN using optimum conditions for each compound.



**Figure 4.** Continuous variation plot for (a) NF and (b) VN. Concentration of solutions of both drugs and the reagent ( $\text{KMnO}_4$ ) was  $1 \times 10^{-2}$  M.

**Table 2.** Values of logarithms of rates and concentrations for NF and VN with alkaline  $\text{KMnO}_4$  using optimum conditions for each compound

Compound	$\log \text{C}[\text{M}]$	$\log \Delta A/\Delta t$
NF	-5.198	-3.799
	-4.896	-3.500
	-4.720	-3.343
	-4.597	-3.259
	-4.499	-3.167
VN	-4.974	-3.741
	-4.772	-3.582
	-4.646	-3.448
	-4.549	-3.348
	-4.469	-3.255
	-4.403	-3.177



Hence,  $K' = 7.46 \text{ sec}^{-1}$  for NF and  $24.13 \text{ sec}^{-1}$  for VN and the reaction is pseudo-first order ( $n = 1$ ) with respect to NF and VN.

## II. Appraisal of Kinetic Methods

The determination of NF and VN under the optimized experimental conditions mentioned above, in which the potassium permanganate and sodium hydroxide concentrations were several hundreds times that of either drug, would result in pseudo-zero conditions with respect to their concentrations and the rate of reaction will be directly proportional to the concentration of NF or VN in a pseudo-first-order rate equation as follows:

$$\text{Rate} = K' \times [C] \dots \dots \dots (3)$$

where  $K'$  is the pseudo-first-order rate constant.

Equation (3) was the basis for several experiments, which were run to obtain concentrations of the investigated drugs using the rate data. Rate constant, fixed-concentration and fixed-time methods<sup>(42)</sup> were tested and the most suitable analytical method was selected taking into account the applicability, sensitivity (i.e. the slope of the calibration graph), correlation coefficient ( $r$ ) and intercept ( $a$ ).

### (I) Rate-constant Method

Graphs of  $\log$  (absorbance) versus time for NF concentrations in the range of  $6.33 \times 10^{-6}$  -  $3.17 \times 10^{-5}$  M (3-15  $\mu\text{g/mL}$ ) and VN concentrations in the range of  $2.26 \times 10^{-5}$  -  $3.95 \times 10^{-5}$  M (8 - 14  $\mu\text{g/mL}$ ) were plotted and all appeared to be rectilinear. Pseudo-first-order rate constants ( $K'$ ) corresponding to different concentrations of the investigated drugs [ $C$ ] were calculated from the slopes multiplied by -2.303. Regression of  $K'$  values versus [ $C$ ] gave the equations:

$$K' = 0.00205 + 18.89 [\text{NF}], r = 0.98364$$

$$K' = -0.00186 + 28.74 [\text{VN}], r = 0.49247$$

The method suffered from poor linearity especially for VN as indicated from its  $r$  value, therefore this method was excluded.

### (II) Fixed-concentration Method

Reaction rates were determined for different concentrations of the investigated drugs. A pre-selected absorbance value was fixed (0.4 for both NF and VN) for different concentrations of the studied drugs, in the range of  $1.27 \times 10^{-5}$  -  $3.17 \times 10^{-5}$  M (6-15  $\mu\text{g/mL}$ ) for NF and  $2.26 \times 10^{-5}$  -  $3.95 \times 10^{-5}$  M (8-14  $\mu\text{g/mL}$ ) for VN and the time required for each concentration to reach the pre-selected absorbance value was measured in seconds. The reciprocal of time ( $1/t$ ) versus drug concentrations was plotted and the following equations were obtained by

linear regression:

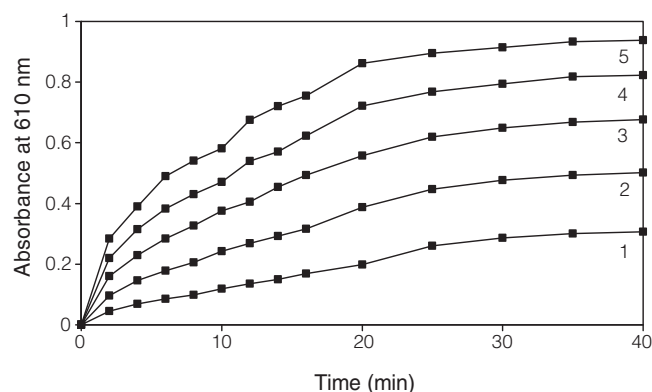
$$1/t = -1.5 \times 10^{-2} + 164.25 [\text{NF}], r = 0.98747$$

$$1/t = -2.6 \times 10^{-2} + 1113.62 [\text{VN}], r = 0.96103$$

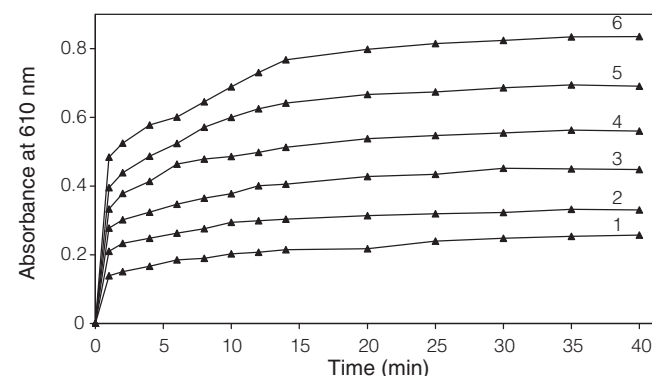
The concentration ranges giving the most satisfactory calibration graphs were limited (6-15  $\mu\text{g/mL}$  for NF and 8-14  $\mu\text{g/mL}$  for VN) with poor linearity and therefore the method is excluded.

### (III) Fixed-time Method

Reaction rates were determined for different concentrations of the investigated drugs. At a pre-selected fixed time, which was accurately determined, the absorbance was measured. Calibration graphs of the absorbance ( $A$ ) versus initial concentration [ $C$ ] were established at different fixed-time intervals of 2-40 min for NF and 1-40 min for VN (Figures 5 and 6) with the regression equations assembled in Table 3. It was found that the slopes increase with time and the most acceptable values for the intercept and the correlation coefficient ( $r$ ) were obtained at a fixed-time of 35 min for NF and 20 min for VN therefore they were chosen as



**Figure 5.** Absorbance versus time graphs for the reaction of NF and alkaline potassium permanganate. Concentrations of NF: (1)  $6.33 \times 10^{-6}$ , (2)  $1.27 \times 10^{-5}$ , (3)  $1.90 \times 10^{-5}$ , (4)  $2.53 \times 10^{-5}$ , (5)  $3.17 \times 10^{-5}$  M.



**Figure 6.** Absorbance versus time graphs for the reaction of VN and alkaline potassium permanganate. Concentrations of VN: (1)  $1.13 \times 10^{-5}$ , (2)  $1.69 \times 10^{-5}$ , (3)  $2.26 \times 10^{-5}$ , (4)  $2.82 \times 10^{-5}$ , (5)  $3.39 \times 10^{-5}$ , (6)  $3.95 \times 10^{-5}$  M.

**Table 3.** Regression equations at different fixed-times for NF in the range  $6.33 \times 10^{-6}$  -  $3.17 \times 10^{-5}$  M and VN in the range  $1.13 \times 10^{-5}$  -  $3.95 \times 10^{-5}$  M with alkaline  $\text{KMnO}_4$  using optimum conditions for each compound

Compound	Time	Regression equation	(r)
NF	2	$A = -0.01943 + 9504.05 C$	0.99929
	4	$A = -0.01354 + 12803.07 C$	0.99971
	6	$A = -0.01976 + 15992.96 C$	0.99971
	8	$A = -0.01196 + 17508.15 C$	0.99974
	10	$A = 0.01194 + 18218.71 C$	0.99832
	12	$A = 0.00022 + 21297.50 C$	0.99998
	14	$A = 0.01161 + 22403.06 C$	0.99909
	16	$A = 0.02745 + 23348.12 C$	0.99825
	20	$A = 0.03274 + 24994.74 C$	0.99844
	25	$A = 0.10503 + 25716.40 C$	0.99769
	30	$A = 0.13035 + 25605.02 C$	0.99757
	35	$A = 0.03954 + 25794.84 C$	0.99741
	40	$A = 0.14524 + 25842.15 C$	0.99734
VN	1	$A = 0.00637 + 11816.06 C$	0.99781
	2	$A = 0.00849 + 12979.56 C$	0.99910
	4	$A = 0.00180 + 14482.97 C$	0.99959
	6	$A = 0.01430 + 15080.14 C$	0.99693
	8	$A = 0.00015 + 16569.02 C$	0.99861
	10	$A = -0.00258 + 17490.14 C$	0.99884
	12	$A = -0.01482 + 18700.25 C$	0.99883
	14	$A = -0.02486 + 19666.37 C$	0.99759
	20	$A = -0.00989 + 20592.51 C$	0.99857
	25	$A = -0.01616 + 20511.65 C$	0.99660
	30	$A = -0.00879 + 20608.64 C$	0.99677
	35	$A = -0.00575 + 20744.71 C$	0.99655
	40	$A = -0.00440 + 20658.66 C$	0.99561

the most suitable time intervals for measurements. The calibration graphs were linear over the concentration ranges of  $6.33 \times 10^{-6}$  -  $3.17 \times 10^{-5}$  M (3-15  $\mu\text{g/mL}$ ) and  $1.13 \times 10^{-5}$  -  $3.95 \times 10^{-5}$  M (4-14  $\mu\text{g/mL}$ ) for NF and VN, respectively.

### III. Analytical Performance of the Proposed Method

#### (I) Concentration Ranges and Calibration Graphs

The analytical parameters, molar absorptivities, linearity ranges and regression equations calculated from calibration graphs along with the standard deviations of the slope ( $S_b$ ), intercept ( $S_a$ ) and the standard deviation of residuals ( $S_{y/x}$ ) are presented in Table 1. The values of the correlation coefficients ( $r$ ) of regression equations indicated good linearity and conformity to Beer's law. The linearity was also evaluated by calculation of percentage relative standard deviation of the slope ( $S_b$  %). It was found to be less than 2% for the developed analytical method. An important statistic indicating the random error in the estimated values of  $y$  is the standard error

**Table 4.** Precision and accuracy for the proposed fixed time kinetic method for the determination of NF and VN

Compound	Nominal Value ( $\mu\text{g/mL}$ )	% Recovery*	RSD (%)
NF	3	100.33	1.16
	6	100.67	1.39
	12	100.08	0.41
VN	4	101.25	1.80
	8	98.00	1.51
	12	102.08	2.65

\*Mean % recovery of five determinations.

of the estimate, standard deviation about regression, or standard deviation of residuals,  $S_{y/x}$ . The smaller the standard error of the estimate, the closer the points are to the straight line.

#### (II) Detection and Quantification Limits

The detection limits were  $8.01 \times 10^{-7}$  and  $1.19 \times 10^{-6}$  M (0.38 and 0.42  $\mu\text{g/mL}$ ) for NF and VN, respectively, while the quantification limits were  $2.67 \times 10^{-6}$  and  $3.97 \times 10^{-6}$  M (1.26 and 1.40  $\mu\text{g/mL}$ ) for NF and VN, respectively. These values were calculated according to the formulae provided by the USP<sup>(43)</sup>.

#### (III) Precision and Accuracy

Five replicate determinations at different concentration levels were carried out to test the precision and accuracy of the proposed fixed time kinetic method. As shown in Table 4, the % recovery and RSD (%) values demonstrated the good repeatability and accuracy of the proposed method.

#### (IV) Robustness

The robustness of the method was demonstrated by the flexibility of the experimental factors that affect the absorbance values. Variation of the volumes added of the both NaOH and  $\text{KMnO}_4$  solutions by  $\pm 5\%$  and variation of the measurement wavelength by  $\pm 2$  nm did not have significant effect on the measured absorbance values.

### IV. Assay of Pharmaceutical Preparations

The fixed-time method was applied to the determination of NF and VN in their commercially available dosage forms. The fixed-time method gave better linearity as indicated from the ( $r$ ) values. In addition it was easier in application than the rate constant method. Direct application of the method on the aqueous extract

**Table 5.** Application of the proposed kinetic method for the determination of NF and VN in their pharmaceutical preparations

Praxilene <sup>®</sup> tablets	Kinetic method	Reference method <sup>b</sup>
%Recovery $\pm$ SD <sup>a</sup>	100.18 $\pm$ 1.84	99.87 $\pm$ 1.99
RSD % <sup>b</sup>	1.84	1.99
t		0.26
F		1.17
Oxybral <sup>®</sup> capsules	Kinetic method	Reference method <sup>c</sup>
%Recovery $\pm$ SD <sup>a</sup>	98.83 $\pm$ 0.74	99.52 $\pm$ 1.25
RSD % <sup>b</sup>	0.75	1.26
t		1.04
F		2.88
Oxybral <sup>®</sup> ampoules	Kinetic method	Reference method <sup>c</sup>
%Recovery $\pm$ SD <sup>a</sup>	99.32 $\pm$ 1.13	100.04 $\pm$ 0.64
RSD % <sup>b</sup>	1.14	0.64
t		1.25
F		3.12

<sup>a</sup>Mean % recovery  $\pm$  SD for five determinations.<sup>b</sup>Measurement of  $A_{\max}$  in saline at 283 nm<sup>(15)</sup>.<sup>c</sup>Measurement of the first derivative of the ratio spectra (<sup>1</sup>DD<sub>293</sub>) of VN over 20  $\mu\text{g mL}^{-1}$  standard VA<sup>(28)</sup>. Theoretical values for t and F at P = 0.05 are 2.31 and 6.39, respectively.

of Praxilene<sup>®</sup> tablets resulted in a positive error in recovery. This error can be attributed to the oxidation of the common excipients usually present in the tablet dosage form (e.g. lactose, starch, etc), therefore, acetonitrile was chosen as a dissolution medium and the reaction was applied on the acetonitrile aliquots. On the other hand, because VN has a limited solubility in water, acetonitrile was also chosen as a solvent for VN. For Oxybral<sup>®</sup> ampoules, an extraction procedure with chloroform has been followed in order to eliminate the interference caused by the antioxidant present in the ampoules which is easily oxidized by  $\text{KMnO}_4$  yielding a positive error in recovery values. The results obtained show good precision and accuracy (Table 5) and they were statistically compared with those obtained by the reference methods<sup>(15,28)</sup>. The student's t-test and the variance-ratio F-test values at 95 % confidence level did not exceed the theoretical values, indicating no significant difference between the proposed and the reference method (Table 5).

### V. Interferences

Interferences due to common excipients and related co-formulated compounds were studied using glucose, lactose, sucrose, starch, ascorbic acid, sodium carboxymethylcellulose, hydroxypropylmethylcellulose, riboflavin and piracetam. Different aliquots of the interfering substance solutions in acetonitrile were added to

**Table 6.** Effect of various foreign species on the determination of NF and VN at the optimum conditions

Species	Tolerance Limit ( $\mu\text{g/mL}$ )	
	NF (10 $\mu\text{g/mL}$ )	VN (10 $\mu\text{g/mL}$ )
Glucose	12.35	12.77
Lactose	14.02	12.35
Sucrose	20.98	29.76
Starch	13.10	16.22
Ascorbic acid	23.44	25.42
Riboflavin	21.43	32.70
Carboxymethylcellulose	16.67	17.24
Hydroxypropylmethylcellulose	12.35	11.49
Piracetam	—	5.45

standard NF and VN solutions, treated and measured at the optimum conditions for each compound. The tolerance limit<sup>(37)</sup> for each of these substances (Concentration of interfering substance causing 3% relative error) was calculated and listed in Table 6.



## CONCLUSIONS

In this study, a simple and sensitive kinetic fixed-time spectrophotometric procedure was developed for the analysis of the two vasodilators: NF and VN. Reviewing the literature exposed that there were no reports for the kinetic spectrophotometric determination of the two drugs. Moreover, nothing was reported concerning the use of colorimetry in the assay of NF, and only single previous report for the UV spectrophotometric estimation of NF<sup>(15)</sup>. On the other hand, very few previous studies were concerned with the spectrophotometric or colorimetric analysis of VN. The simplicity, convenience at low cost and sensitivity of the proposed method are superior or comparable to those of the official non-aqueous titration method<sup>(2)</sup> and several previously published spectrophotometric methods<sup>(15,25-28,31,32)</sup>. Furthermore, the proposed method does not require elaborate treatment or sophisticated experimental setup usually associated with HPLC methods of analysis. The developed method used only a spectrophotometer, which is available in all quality control laboratories, and it involved very simple procedure with readily available chemicals (KMnO<sub>4</sub> and NaOH). The applicability of the developed method was evaluated through the determination of the two drugs in bulk form and in pharmaceutical formulations with good accuracy and precision, therefore it can be considered useful and convenient for the routine and quality control assay of the two drugs.

## REFERENCES

1. Sweetman, S. C. 2007. Martindale: The Complete Drug Reference. 35th pp. 1210, 2186. The Pharmaceutical Press, London.
2. The British Pharmacopoeia 2008. Volumes II and III. pp. 1507-1509, 2918, 2919. Her Majesty's Stationery Office, London, UK.
3. Garrett, E. R. and Barbhuiya, R. 1981. Prediction of stability in pharmaceutical preparations. XVIII: Application of high-pressure liquid chromatographic assays to study of nafronyl stability and bioanalysis. *J. Pharm. Sci.* 70: 39-45.
4. Brodie, R. R., Chasseaud, L. F., Taylor, T., Hunter, J. O. and Ciclitira, P. J. 1979. Determination of naftidrofuryl in the plasma of humans by high-performance liquid chromatography. *J. Chromatogr.* 164: 534-540.
5. Yu, C., Zhang, H., Hong, Y., Cui, G. and Xia, F. 1997. RP-HPLC fluorimetry of naftidrofuryl in serum. *Zhongguo Yiyao Gongye Zazhi* 28: 311-313.
6. Waaler, P. J. and Mueller, B. W. 1992. Solid-phase extraction of naftidrofuryl from human plasma for high-performance liquid chromatography analysis. *Int. J. Pharm.* 87: 223-227.
7. Walmsley, L. M., Wilkinson, P. A., Brodie, R. R. and Chasseaud, L. F. 1985. Determination of naftidrofuryl in human plasma by high-performance liquid chromatography with fluorescence detection. *J. Chromatogr.* 338: 433-437.
8. Beyer, K. H. and Hildebrand, M. 1982. Naftidrofuryl (Dusodril). Possible uses of gas chromatography and high pressure liquid chromatography for analysis. *Dtsch. Apoth. Ztg.* 122: 1709-1712.
9. Cruces-Blanco, C., Segura Carretero, A., Fernandez Sanchez, J. F. and Fernandez Gutierrez, A. 2000. Facile and selective determination of the cerebral vasodilator nafronyl in a commercial formulation by heavy atom induced room temperature phosphorimetry. *J. Pharm. Biomed. Anal.* 23: 845-850.
10. Segura Carretero, A., Cruces Blanco, C., Canabate Diaz, B., Fernandez Sanchez, J. F. and Fernandez Gutierrez, A. 2000. Heavy-atom induced room-temperature phosphorescence: a straightforward methodology for the determination of organic compounds in solution. *Anal. Chim. Acta* 417: 19-30.
11. Murillo Pulgarin, J. A., Alanon Molina, A. and Fernandez Lopez, P. 1999. Phosphorimetric determination of nafronyl in pharmaceutical preparations. *Anal. Chim. Acta* 382: 77-85.
12. Munoz de la Pena, A., Espinosa Mansilla, A., Murillo Pulgarin, J. A., Alanon Molina, A. and Fernandez Lopez, P. 1998. Determination of nafronyl in pharmaceutical preparations by means of stopped-flow micellar-stabilized room temperature phosphorescence. *Analyst* 123: 2285-2290.
13. Ionescu, M. S., Badea, V., Baiulescu, G. E. and Cosofret, V. V. 1986. Nafronyl ion-selective membrane electrodes and their use in pharmaceutical analysis. *Talanta* 33: 101-103.
14. Fernandez-Sanchez, J. F., Segura-Carretero, A., Cruces-Blanco, C. and Fernandez-Gutierrez, A. 2002. Room-temperature luminescence optosensings based on immobilized active principles. Application to nafronyl and naproxen determination in pharmaceutical preparations and biological fluids. *Anal. Chim. Acta* 462: 217-224.
15. Cawood, A. and Marshall, I. W. 1975. Estimation of naftidrofuryl in sodium chloride solution. *J. Hosp. Pharm.* 33: 149-151.
16. Dal Bo, L., Ceriani, G. and Broccali, G. 1992. Determination of vincamine in human plasma by high-performance liquid chromatography with ultraviolet detection. *J. Chromatogr.* 573: 158-162.
17. Smyth, M. R. 1986. Determination of vincamine in plasma by high-performance liquid chromatography with voltammetric detection. *Analyst* 111: 851-852.
18. Dubruc, C., Caqueret, H. and Bianchetti, G. 1981. Determination of vincamine in human plasma using automated high-performance liquid chromatography. *J. Chromatogr.* 204: 335-339.
19. Pietta, P., Rava, A. and Catenacci, E. 1981. High-performance liquid chromatographic determination of vincamine. *J. Chromatogr.* 210: 149-153.

20. Sriewoelan, S. and Bres, J. 1985. Gas chromatographic determination of vincamine using semi-capillary column. *Acta Pharm. Indones.* 10: 46-49.
21. Michotte, Y. and Massart, D. L. 1985. Capillary gas chromatographic determination of vincamine in plasma. *J. Chromatogr.* 344: 367-371.
22. Devaux, P., Godbille, E. and Viennet, R. 1979. Quantitative determination of vincamine in human plasma by gas chromatography-mass spectrometry. *Recent Dev. Mass Spectrom. Biochem. Med.* 2: 191-203.
23. Kinsun, H. and Moulin, M. A. 1977. Gas chromatographic method for the determination of vincamine in blood. *J. Chromatogr.* 144: 123-126.
24. Iven, H. and Siegers, C. P. 1977. Fluorometric assay and pharmacokinetics of vincamine in rats. *Arzneimittelforschung* 27: 1248-1254.
25. El-Saharty, Y. S. I. 2008. Simultaneous determination of piracetam and vincamine by spectrophotometric and high-performance liquid chromatographic methods. *J. AOAC Int.* 91: 311-321.
26. El-Bardicy, M. G., Lotfy, H. M., El-Sayed, M. A. and El-Tarras, M. F. 2008. Smart stability-indicating spectrophotometric methods for determination of binary mixtures without prior separation. *J. AOAC Int.* 91: 299-310.
27. El-Gindy, A., Sallam, S. and Abdel-Salam, R. A. 2006. Liquid chromatography and chemometric-assisted spectrophotometric methods for the simultaneous determination of vincamine with piracetam and diflunisal with naproxen. *Bull. Fac. Pharm. Cairo Univ.* 44: 279-298.
28. Shehata, M. A. M., El Sayed, M. A., El Tarras, M. F. and El Bardicy, M. G. 2005. Stability-indicating methods for determination of vincamine in presence of its degradation product. *J. Pharm. Biomed. Anal.* 38: 72-78.
29. Amato, A., Cavazzutti, G., Gagliardi, L., Profili, M., Zagarese, V., Chimenti, F., Tonelli, D. and Gattavecchia, E. 1983. Determination of vincamine by high-performance liquid chromatography with dual-wavelength ultraviolet detection. *J. Chromatogr.* 270: 387-391.
30. Aboutabl, E. A., El-Azzouny, A. A. and Afifi, M. S. 1998. PMR assay of vincamine and formulations. *Pharm. Acta. Helv.* 73: 193-197.
31. Ganesu, I., Papa, I., Ganesu, A., Bratulescu, G. and Cirtina, D. 2002. Thiocyanatochromic complexes in analytical chemistry. Vincamine determination. *Acta Chim. Slov.* 49: 181-185.
32. Ganesu, I., Mircioiu, C., Papa, I., Ganesu, A., Aldea, V., Chirigiu, L. and Barbu, A. 2001. Thiocyanato-platinum complexes in analytical determination of vincamine. *Farmacia (Bucharest, Romania)* 49: 62-68.
33. Chen, Q., Miao, J. and Ma, J. 1995. Determination of vincamine in tissue culture of *Vinca minor* L. by TLC scanning method. *Zhongguo Yaoke Daxue Xuebao* 26: 350-352.
34. Proksa, B. and Grossmann, E. 1991. High performance liquid chromatographic determination of alkaloids from *Vinca minor* L. *Phytochem. Anal.* 2: 74-76.
35. Crouch, S. R., Cullen, T. F., Scheeline, A. and Kirkor, E. S. 1998. Kinetic determinations and some kinetic aspects of analytical chemistry. *Anal. Chem.* 70: 53R-106R.
36. Hassan, E. M. 2000. Determination of ipratropium bromide in vials using kinetic and first-derivative spectrophotometric methods. *J. Pharm. Biomed. Anal.* 21: 1183-1189.
37. Hassan, E. M. and Belal, F. 2002. Kinetic spectrophotometric determination of nizatidine and ranitidine in pharmaceutical preparations. *J. Pharm. Biomed. Anal.* 27: 31-38.
38. Rahman, N., Ahmad, Y. and Hejaz-Azmi, S. N. 2004. Kinetic spectrophotometric method for the determination of norfloxacin in pharmaceutical formulations. *Eur. J. Pharm. Biopharm.* 57: 359-367.
39. Darwish, I. A. 2005. Kinetic spectrophotometric methods for determination of trimetazidine dihydrochloride. *Anal. Chim. Acta* 551: 222-231.
40. Youssef, R. M., Khamis, E. F., Gazy, A. A., Mahgoub, H. and El-Sayed, M. A. 2006. Assay of buspirone hydrochloride in tablets using kinetic spectrophotometry. *Chin. Pharm. J.* 58: 85-94.
41. Sawyer, D. T., Heineman, W. R. and Beebe, J. M. 1984. *Chemistry Experiments for Instrumental Methods.* pp. 205. John Wiley and Sons, New York.
42. Pérez-Bendito, D. and Silva, M. 1988. *Kinetic Methods in Analytical Chemistry.* pp. 40. John Wiley and Sons, New York.
43. United States Pharmacopeial Convention. 2007. *The United States Pharmacopeia 30th revision. "The Official Compendia of Standards."* 25th ed, volume 1, pp. 682. Inc., Asian Edition, Washington, D.C.